




REVIEW

Non-coding RNA and cholesteatoma

Ivan Jovanovic PhD¹  | Maja Zivkovic PhD¹  | Snezana Jesic MD, PhD^{2,3} | Aleksandra Stankovic PhD¹ ¹VINČA Institute of Nuclear Sciences, National Institute of the Republic of Serbia, University of Belgrade, Belgrade, Serbia²Medical Faculty Belgrade, University of Belgrade, Belgrade, Serbia³Clinic for Otorhinolaryngology and Maxillofacial Surgery, Clinical Centre of Serbia, Belgrade, Serbia**Correspondence**

Aleksandra Stankovic, VINČA Institute of Nuclear Sciences, National Institute of the Republic of Serbia, University of Belgrade, Laboratory for Radiobiology and Molecular Genetics, P.O. Box 522, Belgrade 11001, Serbia.

Email: alexas@vin.bg.ac.rs

Funding information

Ministarstvo Prosvete, Nauke i Tehnološkog Razvoja, Grant/Award Number: 451-03-9/2021-14/200017

Abstract**Objective:** Cholesteatoma is a challenging chronic pathology of the middle ear for which pharmacologic therapies have not been developed yet. Cholesteatoma occurrence depends on the interplay between genetic and environmental factors while master regulators orchestrating disease progression are still unknown. Therefore, in this review, we will discuss the diagnostic and therapeutic potential of non-coding RNAs (ncRNA) as a new class of regulatory molecules.**Methods:** We have comprehensively reviewed all articles investigating ncRNAs, specifically micro RNAs (miRNAs) and long ncRNAs (lncRNA/circRNA) in cholesteatoma tissue.**Results:** Candidate miRNA approaches indicated that miR-21 and let-7a are the major miRNAs involved in cholesteatoma growth, migration, proliferation, bone destruction, and apoptosis. Regulatory potential for the same biological processes was also observed for miR-203a. The NF-κB/miR-802/PTEN regulatory network was in relation to observed miR-21 activity in cholesteatoma as well. High throughput approaches revealed additional ncRNAs implicated in cholesteatoma pathology. Competitive endogenous RNA (ceRNA) analysis highlighted lncRNA/circRNA that could be “endogenous sponge” for miR-21 and let-7a based on the hypothesis that RNA transcripts can communicate with and regulate each other by using shared miRNA response elements.**Conclusion:** In this review, we summarize the discoveries and role of ncRNA in major pathways in cholesteatoma and highlight the potential of miRNA-based therapeutics in the treatment of cholesteatoma.**Level of Evidence:** NA.**KEYWORDS**

cholesteatoma, long noncoding RNA, micro RNA, noncoding RNA, RNA interaction

1 | MOLECULAR BASIS OF THE CHOLESTEATOMA DEVELOPMENTCholesteatoma development may depend on the interplay between genetic and environmental factors,¹ however, the molecular mechanismsunderlying cholesteatoma pathogenesis remain undefined. Cholesteatoma investigations have progressed from evaluating individual candidate genes to genome-wide studies to elucidate molecular mechanisms of development on a genomic scale.²⁻⁵ Despite that numerous processes and pathways have been determined to harbor dysregulated genes identified in

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2022 The Authors. *Laryngoscope Investigative Otolaryngology* published by Wiley Periodicals LLC on behalf of The Triological Society.

cholesteatoma,²⁻⁶ no discrete pharmacological targets have been identified yet. To identify master regulators of these pathways and provide pharmacologic targets for medical management of cholesteatoma, we need to look beyond the protein-coding genes and into the universe of noncoding RNA (ncRNA) molecules (Table 1).

2 | THE EXPANDING UNIVERSE OF ncRNAs

Only 1%-2% of the human genome is transcribed in coding RNAs, able to encode a sequence of amino acids in proteins.¹⁵ The RNA that does not code for proteins are called ncRNA. In this narrative review, we will focus on the two ncRNA subgroups, short ncRNAs and long ncRNAs^{16,17} because of their diagnostic and therapeutic potential.

2.1 | Small ncRNA

The dominant class of small ncRNAs, microRNAs (miRNAs), are around 20 nucleotides in size. miRNAs orchestrate gene expression of almost every biological process¹⁸⁻²⁰ and consequently are associated with various diseases.²¹⁻²³ miRNAs are negative regulators of gene expression acting through induction of mRNA degradation or inhibition of its translation (Figure 1A).²⁴ Different miRNAs could share the same target mRNA while single miRNA could target multiple mRNAs.¹⁸ These properties of miRNA molecules meet most of the required criteria for an ideal biomarker,²⁵ while miRNA mimics and antagomiRNA are considered as promising therapeutics.²⁶ As miRNA dysregulation has been

observed in cholesteatoma, in this review, we will highlight the subsequent effects on cellular mechanisms and possible medical applications.

2.2 | Long ncRNA

lncRNAs show great capacity for gene expression regulation at both transcriptional and posttranscriptional levels through the sequence- and structure-specific mechanism.^{27,28} Competitive endogenous RNA (ceRNA) hypothesis states that all types of RNA transcripts can communicate with and regulate each other by using shared miRNA response elements (MREs).²⁹ lncRNA can act as miRNA decoy capturing active miRNAs, buffering that way regulatory activity of those miRNA on their target mRNAs, which share the same MREs.^{29,30} Subclass of ncRNAs are circular RNAs (circ-RNA),³¹ covalently closed continuous loops, highly conserved and tissue-specific.³²⁻³⁴ circRNAs have been reported to harbor multiple miRNA binding sites, which seems to be a typical feature of this class of RNA molecules.^{35,36} Interaction between the miRNA and lncRNA/circRNA represents a complex interaction system (Figure 1B). This mechanism of gene expression regulation, important in all aspects of physiology and disease,³⁷⁻³⁹ will be discussed in the context of cholesteatoma molecular pathology research.

3 | LITERATURE SEARCH AND INCLUSION CRITERIA

Literature search was carried out on Pubmed.gov database using different combinations of keywords: (cholesteatoma) AND ((miRNA) OR

TABLE 1 miRNAs associated with cholesteatoma

miRNAs associated with cholesteatoma	Regulated direction of expression	miRNA expression validation	Regulatory mechanism in cholesteatoma	Cellular function	Proposed ceRNA interaction in cholesteatoma	References
miR-21	Upregulated in cholesteatoma versus normal skin	qRT-PCR	Downregulation of PTEN and PDCD4	Increased proliferation of keratinocytes	lncRNA-uc001kfc.1 circRNA-102747	[7-10]
let-7a	Upregulated in cholesteatoma versus normal skin	qRT-PCR	Downregulation of HMGA2	Decreased proliferation of keratinocytes	circRNA-101458	[8,10,11]
miR-802	Upregulated upon NF-κB activation	miRNA transfection qRT-PCR	Downregulation of PTEN	Increased proliferation of keratinocytes in vitro	—	[12]
miR-203a	Downregulated in cholesteatoma versus normal skin	qRT-PCR	Upregulated Bmi1 and subsequent increase of p-Akt level	Increased proliferation, migration, and antiapoptotic abilities	—	[13]
miR-16-1-3p ^a	Upregulated in cholesteatoma versus normal skin	Microarray qRT-PCR	Regulation of PI3K/Akt signaling pathway	Hyper-proliferation of cholesteatoma	—	[14]
miR-10a-5p ^a	Downregulated in cholesteatoma versus normal skin	Microarray qRT-PCR	Regulation of PI3K/Akt signaling pathway	Hyper-proliferation of cholesteatoma	—	[14]

^aOnly qRT-PCR validated miRNAs identified in high throughput analysis have been presented.

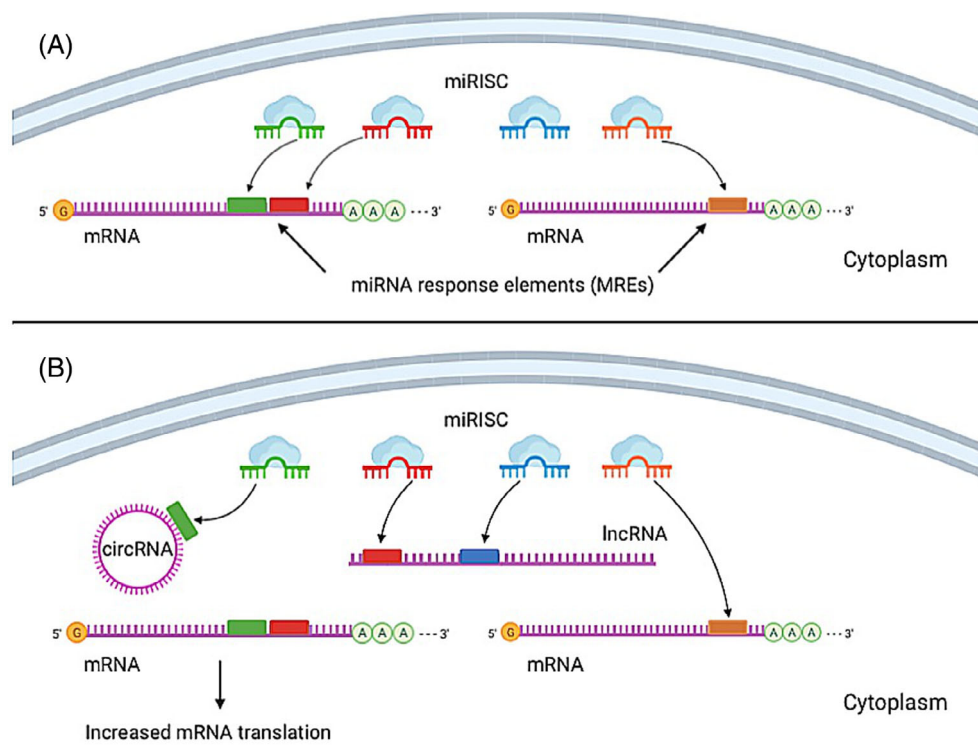


FIGURE 1 Noncoding RNA interplay in regulation of the mRNA expression. The figure depicts the mechanism of gene expression regulation in the absence and presence of competing endogenous RNA (ceRNA) interactions with miRNAs. (A) In the absence of ceRNA (circRNA and lncRNA) various miRNAs incorporated in RNA-induced silencing complex (miRISC) induce degradation or repression of translation of target mRNA; (B) When ceRNAs (circRNA and lncRNA) harboring the same miRNA response elements (MREs) as the target mRNAs are present, they complementary sequester miRISCs (green, red, or blue) and consequently lower cellular levels of free miRISCs, which leads to increased translation; Cellular levels of miRISC (orange) remains unchanged since lncRNAs do not harbor the same MREs as the target mRNA, thus not affecting the miRNA-mRNA regulation, which results in unchanged, miRNA-induced target mRNA degradation, or repression of translation

(miR); (cholesteatoma) AND ((long noncoding RNA) OR (lncRNA) OR (ceRNA)); (cholesteatoma) AND (miRNA) AND (microarray) ending with August 2020. Studies in which ncRNA profiling was performed in cholesteatoma tissue from adult and pediatric patients were included in the review. Studies investigating ncRNA expression in samples other than cholesteatoma tissue from patients with cholesteatoma were excluded. Based on the defined search and inclusion criteria, eight studies represented the basis of this review. Although one additional study has passed the search criteria it was not included in this review due to the incompatibility with inclusion criteria.

4 | ARE miR-21 AND LET-7 BALANCING CHOLESTEATOMA BETWEEN BENIGN AND INVASIVE NATURE?

The first two miRNAs assigned to be important in cholesteatoma were miR-21 and let-7 as well as their interplay.^{7,8,11} Both miRNAs have been found upregulated in cholesteatoma compared to normal skin^{7,8} with a more pronounced difference in pediatric samples.⁸ At the time the studies have been performed, miR-21 was known to be an onco-miR due to its role in tumorigenesis through regulation of potent tumor suppressors such as PTEN and PDCD4.^{40,41} The downregulation of

PTEN in cholesteatoma compared with levels in normal skin inversely correlates with p-Akt levels.⁴² PI3K-Akt pathway is important for the induction of cell proliferation and terminal differentiation,⁴³ and thus was proposed as a possible mechanism of development and progression of cholesteatoma through downregulation of PTEN.⁴² In addition to PTEN, PDCD4 was also shown to suppress benign and malignant skin tumor formation and progression.⁴⁴ This critical regulator of apoptosis, which inhibits the procaspase-3 mRNA translation, is shown to be dependent on miRNA regulation under apoptotic stimuli.⁴⁵ It was indeed confirmed by western blot that downregulation of PTEN and PDCD4 correlates with upregulation of miR-21, making a significantly greater reduction in PTEN and PDCD4 protein levels in pediatric versus adult cholesteatoma.⁸

Additionally, a similar profile of expression changes both between cholesteatoma and normal skin and between adult and pediatric cholesteatoma was observed for let-7a.⁸ By investigating protein levels of its target HMGA2, it was shown that its levels are reduced in cholesteatomas, especially in pediatric cholesteatomas.⁸ This small, nonhistone chromatin-associated protein has no intrinsic regulatory activity on gene expression. However, its capability to alter chromatin architecture could influence gene transcription through the influence on the assembly of multiprotein complexes of transcriptional factors.⁴⁶ It was reported that in vitro disruption of HMGA2 suppression by let-7 miRNA enhances

oncogenic transformation.^{47,48} Eventual inhibition of HMGA2 by let-7a upregulation in cholesteatoma may lead to increased keratinocyte apoptosis and a reduction in the proliferation of cholesteatoma cells.

Based on findings of miR-21 and let-7a expression in cholesteatoma and joint regulation of their targets, a balancing mechanism has been proposed, perpetuating the growth and invasiveness of cholesteatoma by PTEN and PDCD4 downregulation but keeping it in a benign stage through HMGA2 inhibition.⁸ To explain the interplay of the two miRNAs in cholesteatoma it was shown that let-7a transfected mimics inhibited the growth, migration, and invasion of cholesteatoma keratinocytes *in vitro*.¹¹ It was suggested that the observed effect in cholesteatoma keratinocytes could be explained by the mechanism through which let-7a downregulates miR-21, causing subsequent regulation of its targets.¹¹ However, knowing that both miR-21 and let-7 were upregulated in cholesteatoma *ex vivo*, implies that the sole interaction between the induced expression of let-7 and downregulation of miR-21 was not a sufficient mechanism. Additional mechanistic studies are needed to confirm this interesting hypothesis aiming to tackle the paradoxical nature of cholesteatoma.

5 | NF- κ B/miR-802/PTEN REGULATORY NETWORK BEYOND miR-21 IN CHOLESTEATOMA

As previously mentioned, multiple miRNAs could share the same target mRNA¹⁸ making the additive fine-tuning of gene expression. This effect was also found to be important in cholesteatoma, by describing additional PTEN regulation, beyond miR-21, through the first time revealed NF- κ B/miR-802/PTEN regulatory network.¹² The increased mRNA levels of proinflammatory cytokines TNF- α , IL-1 β , and IL-6 and increased phosphorylation of NF- κ B subunit P65 in the clinical samples of cholesteatoma tissues compared to retroauricular skin have been found.¹² Through comprehensive analysis integrating bioinformatics and *in vitro* manipulation of miR-802 in primary keratinocytes, it was demonstrated that activated NF- κ B increase expression levels of miR-802, which subsequently promotes cell proliferation through regulation of PTEN.¹² The observed mechanism was suggested to be important in cholesteatoma development and progression as well.¹² However, there are no studies investigating miR-802 expression in cholesteatoma tissue, while the single high-throughput study performing miRNA expression profiling in cholesteatoma compared to normal skin to date has not reported differential expression of miR-802.¹⁴ These discrepancies warrant further *ex vivo* validation in cholesteatoma tissue on a larger number of samples to confirm the *in vitro* observations especially by taking into account the complex perimatrix-matrix molecular interplay which has been suggested to play a major role in cholesteatoma.⁶

6 | miR-203 IN CHOLESTEATOMA PATHOLOGY

The next miRNA investigated in cholesteatoma pathology, miR-203a¹³ is specific to epithelial tissue by affecting the growth,

differentiation, and physiology of keratinocytes; and is an important contributor to skin development.^{49–51} Additionally, studies investigating miR-203a in various cancer pathologies have suggested its cancer-suppressive potential.^{52–55} Also delay of chronic wound healing is linked with over-expression of miR-203 in diabetes mellitus.⁵⁶ miR-203a is significantly lower in cholesteatoma than in normal retroauricular skin, while its bioinformatically predicted target Bmi1 showed correlated upregulation on the protein level. Using luciferase assay on HaCaT cell model, the miR-203/Bmi1 interaction has been confirmed, while the same cells displayed hyperproliferation, a low rate of apoptosis, and abnormal migration in low levels of miR-203.¹³ It has been described that Bmi1 increases the level of p-Akt, which is considered as one of the mechanisms of tumor cell proliferation, migration, and antiapoptotic abilities.^{57–59} p-Akt was also linked with the development of cholesteatoma.^{60,61} The upregulation of EGFR/Akt/NF- κ B/cyclinD1 survival signaling pathway in cholesteatoma epithelium compared to normal skin was suggested to be involved in cellular hyperplasia of cholesteatoma,⁶⁰ while its activity in PI3K/Akt/PKB survival signaling may be an additional factor of early keratinocyte differentiation arrest.⁶¹ Further research of the upstream mechanisms responsible for miR-203a downregulation in cholesteatoma remains to be elucidated.

7 | HIGH-THROUGHPUT miRNA EXPRESSION PROFILING IN CHOLESTEATOMA

miRNA microarray technique for miRNA expression profiling identified 44 upregulated and 175 downregulated miRNAs in acquired middle ear cholesteatoma compared to normal skin.¹⁴ The bioinformatic analysis of 19 candidate miRNAs suggested that these miRNAs might be important factors in the etiopathogenesis of middle ear cholesteatoma, by regulating genes involved in cell proliferation, apoptosis, cell cycle, differentiation, bone resorption, and remodeling.¹⁴ Subsequent qRT-PCR validation of miRNAs of particular interest has confirmed the upregulation of miR-21-3p and miRNA-16-1-3p while miRNA-10a-5p levels were decreased in cholesteatoma versus normal skin.¹⁴ However, the discrepancy in the expression of miRNA-584-5p and miRNA-338-5p between qRT-PCR validation and microarray analysis warrants the need for further studies with a larger number of samples. Although the study has a limitation in sample size, the robustness of the high throughput methodology makes acquired data valuable information and a good starting point for the development of novel research hypotheses involving novel miRNA candidates and mechanisms of their activity in cholesteatoma development.

8 | REGULATION OF miR-21 AND LET-7 EFFECTS BY lncRNA IN CHOLESTEATOMA

ceRNA research in cholesteatoma proposed “endogenous sponges” for miR-21-3p and let-7a-3p.^{9,10} Using network analysis of lncRNA/miRNA/mRNA interactions, the authors have discovered lncRNA-

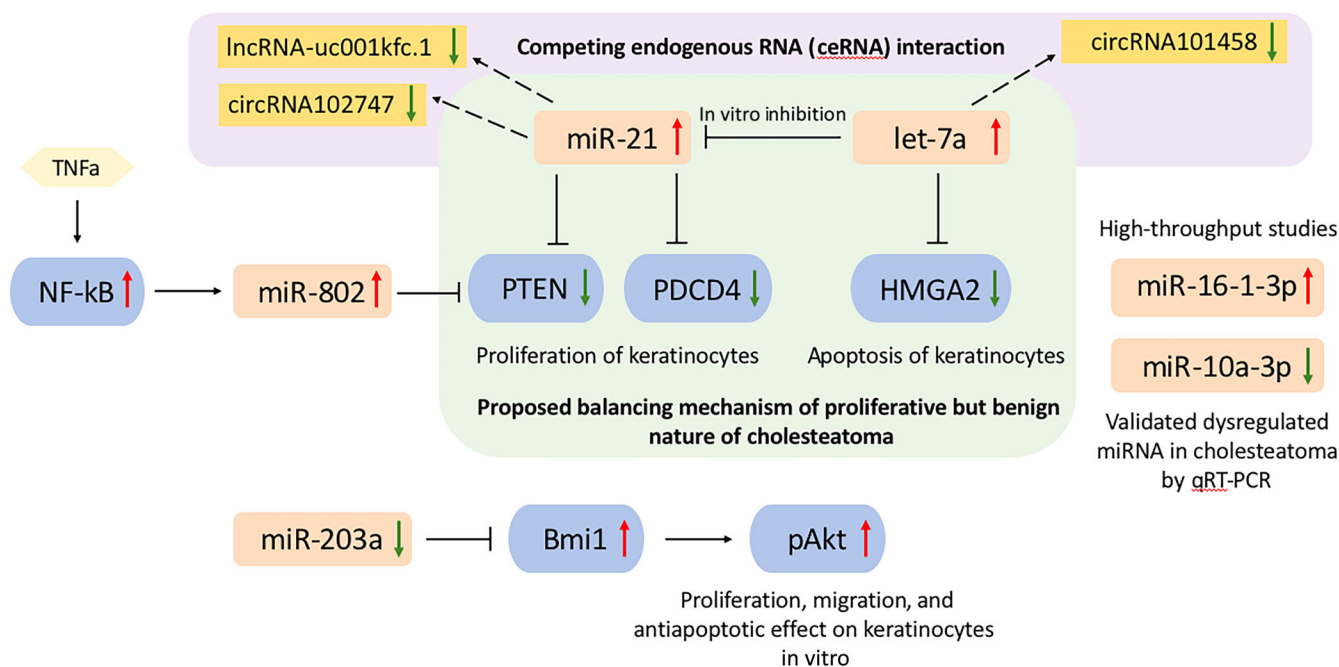


FIGURE 2 The network of described ncRNA activity in cholesteatoma pathogenesis. The Network depicts interplay of ncRNAs in cholesteatoma and regulation of target genes associated with cholesteatoma pathology. Orange boxes emphasize miRNAs which are potential biomarkers and targets for miRNA therapeutics in cholesteatoma. Yellow boxes emphasize lncRNAs and circRNAs as potential regulators of miRNA actions on gene expression. Blue boxes represent target genes implicated in cholesteatoma, related to presented ncRNA interplay

uc001kfc.1 as a possible regulator of miR-21 effects in cholesteatoma and hypothesized that this lncRNA molecule could be a potential drug target for the treatment of cholesteatoma.⁹ The same group aimed to investigate the ceRNA hypothesis in cholesteatoma with circRNAs.¹⁰ The most notable observation was that circRNA-102747 and circRNA-101458, downregulated in cholesteatoma compared to normal skin, interact with miR-21-3p and let-7a-3p. However, other interactions with targeting miRNAs, beyond miR-21 and let-7a, detected in both studies^{9,10} should not be neglected in upcoming cholesteatoma research.

9 | PERSPECTIVES

The important role of ncRNAs in cholesteatoma pathology and the contemporary scarcity of the experimental results in the field could be easily deduced from the thorough analysis of available literature encompassed by the current review. We have integrated and highlighted the progress of the previous research of ncRNAs in cholesteatoma (Table 1, Figure 2). The key ncRNAs and target genes are joined in Figure 2 to depict regulation of ncRNAs in cholesteatoma, their interplay, and putative modulation of target genes associated with pathology.

Based on our literature review we can conclude that miR-21 and let-7a are the two most highlighted ncRNAs dysregulated in cholesteatoma pathology (Figure 2). It is widely recognized that ncRNAs exert many biological properties that can make them noteworthy biomarkers for disease follow-up and minimally invasive therapeutics.⁶² Therapeutics based on blockade of miRNA in different

pathologies are reported recently. Beneficial effects on miR-21 expression inhibition have been observed with miR-21 antagonist in hypertrophic scarring of the skin⁶³ and inhibition of acute tissue injury in LPS-treated human pulmonary alveolar epithelial cells (HPAEPiC).⁶⁴ Even more, a personalized nanomedicine approach with functional nanoparticles was proposed to target both, miR-21 and target genes using the same therapeutic.⁶⁵ For the let-7, clinical trials are already ongoing for the evaluation of the therapeutic potential of this nc-RNA in various diseases such as obesity, diabetes, and cancer.⁶⁶ Although the recently developed analytical models indicate time requirement before many miRNA products reach the market⁶⁷ the progress which has been made in the past few years envisages a promising future.

CONFLICT OF INTEREST

The authors declare no potential conflict of interest.

ORCID

Ivan Jovanovic <https://orcid.org/0000-0001-7932-9463>

Maja Zivkovic <https://orcid.org/0000-0002-0447-6626>

Aleksandra Stankovic <https://orcid.org/0000-0002-1050-5913>

REFERENCES

- Jennings BA, Prinsley P, Philpott C, Willis G, Bhutta MF. The genetics of cholesteatoma. A systematic review using narrative synthesis. *Clin Otolaryngol*. 2018;43(1):55-67. doi:10.1111/coa.12900
- Kwon KH, Kim SJ, Kim HJ, Jung HH. Analysis of gene expression profiles in cholesteatoma using oligonucleotide microarray. *Acta Otolaryngol*. 2006;126(7):691-697. doi:10.1080/00016480500475633
- Klenke C, Janowski S, Borck D, et al. Identification of novel cholesteatoma-related gene expression signatures using full-genome

- microarrays. Chuang EY, ed. *PLoS One*. 2012;7(12):e52718. doi:10.1371/journal.pone.0052718
4. Macias JD, Gerkin RD, Locke D, Macias MP. Differential gene expression in cholesteatoma by DNA chip analysis. *Laryngoscope*. 2013;123: S1-S21. doi:10.1002/lary.24176
 5. Tokuriki M, Noda I, Saito T, et al. Gene expression analysis of human middle ear cholesteatoma using complementary DNA arrays. *Laryngoscope*. 2003;113(5):808-814. doi:10.1097/00005537-200305000-00008
 6. Jovanovic I, Zivkovic M, Djuric T, Stojkovic L, Jesic S, Stankovic A. Perimatrix of middle ear cholesteatoma: a granulation tissue with a specific transcriptomic signature. *Laryngoscope*. 2020;130(4):E220-E227. doi:10.1002/lary.28084
 7. Friedland DR, Eernisse R, Erbe C, Gupta N, Cioffi JA. Cholesteatoma growth and proliferation: posttranscriptional regulation by microRNA-21. *Otol Neurotol*. 2009;30(7):998-1005. doi:10.1097/MAO.0b013e3181b4e91f
 8. Chen X, Qin Z. Post-transcriptional regulation by MicroRNA-21 and *let-7a* MicroRNA in paediatric cholesteatoma. *J Int Med Res*. 2011;39(6):2110-2118. doi:10.1177/147323001103900607
 9. Gao J, Tang Q, Zhu X, et al. Long noncoding RNAs show differential expression profiles and display ceRNA potential in cholesteatoma pathogenesis. *Oncol Rep*. 2018;39(5):2091-2100. doi:10.3892/or.2018.6320
 10. Gao J, Tang Q, Xue R, et al. Comprehensive circular RNA expression profiling with associated ceRNA network reveals their therapeutic potential in cholesteatoma. *Oncol Rep*. 2020;43(4):1234-1244. doi:10.3892/or.2020.7501
 11. ZHANG W, CHEN X, QIN Z. MicroRNA *let-7a* suppresses the growth and invasion of cholesteatoma keratinocytes. *Mol Med Rep*. 2015;11(3):2097-2103. doi:10.3892/mmr.2014.2971
 12. Li N, Qin Z-B. Inflammation-induced miR-802 promotes cell proliferation in cholesteatoma. *Biotechnol Lett*. 2014;36(9):1753-1759. doi:10.1007/s10529-014-1545-y
 13. Zang J, Hui L, Yang N, Yang B, Jiang X. Downregulation of MiR-203a inhibits Bmi1 and promotes growth and proliferation of keratinocytes in cholesteatoma. *Int J Med Sci*. 2018;15(5):447-455. doi:10.7150/ijms.22410
 14. Xie S, Liu X, Pan Z, et al. Microarray analysis of differentially-expressed microRNAs in acquired middle ear cholesteatoma. *Int J Med Sci*. 2018;15(13):1547-1554. doi:10.7150/IJMS.26329
 15. ENCODE project consortium. An integrated encyclopedia of DNA elements in the human genome. *Nature*. 2012;489(7414):57-74. doi:10.1038/nature11247
 16. Mattick JS, Rinn JL. Discovery and annotation of long noncoding RNAs. *Nat Struct Mol Biol*. 2015;22(1):5-7. doi:10.1038/nsmb.2942
 17. Morris KV, Mattick JS. The rise of regulatory RNA. *Nat Rev Genet*. 2014;15(6):423-437. doi:10.1038/nrg3722
 18. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell*. 2009;136(2):215-233. doi:10.1016/j.cell.2009.01.002
 19. Krol J, Loedige I, Filipowicz W. The widespread regulation of microRNA biogenesis, function and decay. *Nat Rev Genet*. 2010;11(9):597-610. doi:10.1038/nrg2843
 20. Carthew RW, Sontheimer EJ. Origins and mechanisms of miRNAs and siRNAs. *Cell*. 2009;136(4):642-655. doi:10.1016/j.cell.2009.01.035
 21. Zeljic K, Jovanovic I, Jovanovic J, Magic Z, Stankovic A, Supic G. MicroRNA meta-signature of oral cancer: evidence from a meta-analysis. *Ups J Med Sci*. 2018;123(1):43-49. doi:10.1080/03009734.2018.1439551
 22. Jovanović I, Zivković M, Jovanović J, Djurić T, Stanković A. The co-inertia approach in identification of specific microRNA in early and advanced atherosclerosis plaque. *Med Hypotheses*. 2014;83(1):11-15. doi:10.1016/j.mehy.2014.04.019
 23. Jovanovic I, Zivkovic M, Kostic M, et al. Transcriptome-wide based identification of miRs in congenital anomalies of the kidney and urinary tract (CAKUT) in children: the significant upregulation of tissue miR-144 expression. *J Transl Med*. 2016;14(1):193. doi:10.1186/s12967-016-0955-0
 24. Kim VN, Nam JW. Genomics of microRNA. *Trends Genet*. 2006;22(3):165-173. doi:10.1016/j.tig.2006.01.003
 25. Condrat CE, Thompson DC, Barbu MG, et al. miRNAs as biomarkers in disease: latest findings regarding their role in diagnosis and prognosis. *Cell*. 2020;9(2):276. doi:10.3390/cells9020276
 26. Lu Q, Wu R, Zhao M, Garcia-Gomez A, Ballestar E. miRNAs as therapeutic targets in inflammatory disease. *Trends Pharmacol Sci*. 2019;40(11):853-865. doi:10.1016/j.tips.2019.09.007
 27. Mercer TR, Dinger ME, Mattick JS. Long non-coding RNAs: insights into functions. *Nat Rev Genet*. 2009;10(3):155-159. doi:10.1038/nrg2521
 28. Wilusz JE, Sunwoo H, Spector DL. Long noncoding RNAs: functional surprises from the RNA world. *Genes Dev*. 2009;23(13):1494-1504. doi:10.1101/gad.1800909
 29. Salmena L, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell*. 2011;146(3):353-358. doi:10.1016/j.cell.2011.07.014
 30. Bak RO, Mikkelsen JG. miRNA sponges: soaking up miRNAs for regulation of gene expression. *Wiley Interdiscip Rev RNA*. 2014;5(3):317-333. doi:10.1002/wrna.1213
 31. Hansen TB, Wiklund ED, Bramsen JB, et al. miRNA-dependent gene silencing involving Ago2-mediated cleavage of a circular antisense RNA. *EMBO J*. 2011;30(21):4414-4422. doi:10.1038/emboj.2011.359
 32. Jeck WR, Sorrentino JA, Wang K, et al. Circular RNAs are abundant, conserved, and associated with ALU repeats. *RNA*. 2013;19(2):141-157. doi:10.1261/rna.035667.112
 33. Jeck WR, Sharpless NE. Detecting and characterizing circular RNAs. *Nat Biotechnol*. 2014;32(5):453-461. doi:10.1038/nbt.2890
 34. Salzman J, Chen RE, Olsen MN, Wang PL, Brown PO. Cell-type specific features of circular RNA expression. *PLoS Genet*. 2013;9(9):e1003777. doi:10.1371/journal.pgen.1003777
 35. Hansen TB, Jensen TI, Clausen BH, et al. Natural RNA circles function as efficient microRNA sponges. *Nature*. 2013;495(7441):384-388. doi:10.1038/nature11993
 36. Memczak S, Jens M, Elefsinioti A, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature*. 2013;495(7441):333-338. doi:10.1038/nature11928
 37. Zhang X, Zhou Y, Chen S, Li W, Chen W, Gu W. LncRNA MACC1-AS1 sponges multiple miRNAs and RNA-binding protein PTBP1. *Oncogenesis*. 2019;8(12):73. doi:10.1038/s41389-019-0182-7
 38. Li R, Fang L, Pu Q, et al. MEG3-4 is a miRNA decoy that regulates IL-1 β abundance to initiate and then limit inflammation to prevent sepsis during lung infection. *Sci Signal*. 2018;11(536):eaao2387. doi:10.1126/scisignal.aao2387
 39. Shan Y, Ma J, Pan Y, Hu J, Liu B, Jia L. LncRNA SNHG7 sponges miR-216b to promote proliferation and liver metastasis of colorectal cancer through upregulating GALNT1. *Cell Death Dis*. 2018;9(7):722. doi:10.1038/s41419-018-0759-7
 40. Asangani IA, Rasheed SAK, Nikolova DA, et al. MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pdc4 and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene*. 2008;27(15):2128-2136. doi:10.1038/sj.onc.1210856
 41. Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology*. 2007;133(2):647-658. doi:10.1053/j.gastro.2007.05.022
 42. Yune TY, Byun JY. Expression of PTEN and phosphorylated Akt in human cholesteatoma epithelium. *Acta Otolaryngol*. 2009;129(5):501-506. doi:10.1080/00016480802258802
 43. Murayama K, Kimura T, Tarutani M, et al. Akt activation induces epidermal hyperplasia and proliferation of epidermal progenitors. *Oncogene*. 2007;26(33):4882-4888. doi:10.1038/sj.onc.1210274

44. Jansen AP, Camalier CE, Colburn NH. Epidermal expression of the translation inhibitor programmed cell death 4 suppresses tumorigenesis. *Cancer Res.* 2005;65(14):6034-6041. doi:10.1158/0008-5472.CAN-04-2119
45. Eto K, Goto S, Nakashima W, Ura Y, Abe SI. Loss of programmed cell death 4 induces apoptosis by promoting the translation of procaspase-3 mRNA. *Cell Death Differ.* 2012;19(4):573-581. doi:10.1038/cdd.2011.126
46. Fedele M, Battista S, Kenyon L, et al. Overexpression of the HMGA2 gene in transgenic mice leads to the onset of pituitary adenomas. *Oncogene.* 2002;21(20):3190-3198. doi:10.1038/sj.onc.1205428
47. Mayr C, Hemann MT, Bartel DP. Disrupting the pairing between let-7 and Hmga2 enhances oncogenic transformation. *Science.* 2007;315(5818):1576-1579. doi:10.1126/science.1137999
48. Park S-M, Shell S, Radjabi AR, et al. Let-7 prevents early cancer progression by suppressing expression of the embryonic gene HMGA2. *Cell Cycle.* 2007;6(21):2585-2590. doi:10.4161/cc.6.21.4845
49. Yi R, Poy MN, Stoffel M, Fuchs E. A skin microRNA promotes differentiation by repressing "stemness". *Nature.* 2008;452(7184):225-229. doi:10.1038/nature06642
50. Wei T, Orfanidis K, Xu N, et al. The expression of microRNA-203 during human skin morphogenesis. *Exp Dermatol.* 2010;19(9):854-856. doi:10.1111/j.1600-0625.2010.01118.x
51. Nissan X, Denis JA, Saidani M, Lemaitre G, Peschanski M, Baldeschi C. miR-203 modulates epithelial differentiation of human embryonic stem cells towards epidermal stratification. *Dev Biol.* 2011;356(2):506-515. doi:10.1016/j.ydbio.2011.06.004
52. Saini S, Majid S, Yamamura S, et al. Regulatory role of mir-203 in prostate cancer progression and metastasis. *Clin Cancer Res.* 2011;17(16):5287-5298. doi:10.1158/1078-0432.CCR-10-2619
53. Okumura T, Shimada Y, Moriyama M, et al. MicroRNA-203 inhibits the progression of esophageal squamous cell carcinoma with restored epithelial tissue architecture in vivo. *Int J Oncol.* 2014;44(6):1923-1932. doi:10.3892/ijo.2014.2365
54. Chi Y, Jin Q, Liu X, et al. miR-203 inhibits cell proliferation, invasion, and migration of non-small-cell lung cancer by downregulating RGS17. *Cancer Sci.* 2017;108(12):2366-2372. doi:10.1111/cas.13401
55. Chen L-Z, Ding Z, Zhang Y, He S-T, Wang X-H. MiR-203 overexpression promotes prostate cancer cell apoptosis and reduces ADM resistance. *Eur Rev Med Pharmacol Sci.* 2018;22(12):3734-3741. doi:10.26355/eurrev_201806_15253
56. Liu J, Shu B, Zhou Z, et al. Involvement of miRNA203 in the proliferation of epidermal stem cells during the process of DM chronic wound healing through Wnt signal pathways. *Stem Cell Res Ther.* 2020;11(1):348. doi:10.1186/s13287-020-01829-x
57. Wang M-C, Jiao M, Wu T, et al. Polycomb complex protein BMI-1 promotes invasion and metastasis of pancreatic cancer stem cells by activating PI3K/AKT signaling, an ex vivo, in vitro, and in vivo study. *Oncotarget.* 2016;7(8):9586-9599. doi:10.18632/oncotarget.7078
58. Liu Y-L, Jiang S-X, Yang Y-M, Xu H, Liu J-L, Wang X-S. USP22 acts as an oncogene by the activation of BMI-1-mediated INK4a/ARF pathway and Akt pathway. *Cell Biochem Biophys.* 2012;62(1):229-235. doi:10.1007/s12013-011-9287-0
59. Xu Z, Liu H, Lv X, Liu Y, Li S, Li H. Knockdown of the Bmi-1 oncogene inhibits cell proliferation and induces cell apoptosis and is involved in the decrease of Akt phosphorylation in the human breast carcinoma cell line MCF-7. *Oncol Rep.* 2011;25(2):409-418. doi:10.3892/or.2010.1078
60. Liu W, Yin T, Ren J, et al. Activation of the EGFR/Akt/NF-κB/cyclinD1 survival signaling pathway in human cholesteatoma epithelium. *Eur Arch Otorhinolaryngol.* 2014;271(2):265-273. doi:10.1007/s00405-013-2403-6
61. Huisman MA, De Heer E, Grote JJ. Survival signaling and terminal differentiation in cholesteatoma epithelium. *Acta Otolaryngol.* 2007;127(4):424-429. doi:10.1080/00016480600868430
62. Rupaimoole R, Slack FJ. MicroRNA therapeutics: towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov.* 2017;16(3):203-221. doi:10.1038/nrd.2016.246
63. Guo L, Xu K, Yan H, et al. MicroRNA expression signature and the therapeutic effect of the microRNA-21 antagomir in hypertrophic scarring. *Mol Med Rep.* 2017;15(3):1211-1221. doi:10.3892/mmr.2017.6104
64. Ge J, Yao Y, Jia H, Li P, Sun W. Inhibition of miR-21 ameliorates LPS-induced acute lung injury through increasing B cell lymphoma-2 expression. *Innate Immun.* 2020;26(8):693-702. doi:10.1177/1753425920942574
65. Gilles ME, Hao L, Brown K, Lim J, Bhatia SN, Slack FJ. Tumor penetrating nanomedicine targeting both an oncomiR and an oncogene in pancreatic cancer. *Oncotarget.* 2019;10(51):5349-5358. doi:10.18632/oncotarget.27160
66. Gilles ME, Slack FJ. Let-7 microRNA as a potential therapeutic target with implications for immunotherapy. *Expert Opin Ther Targets.* 2018;22(11):929-939. doi:10.1080/14728222.2018.1535594
67. Beierlein JM, McNamee LM, Ledley FD. As technologies for nucleotide therapeutics mature, products emerge. *Mol Ther - Nucleic Acids.* 2017;9:379-386. doi:10.1016/j.omtn.2017.10.017

How to cite this article: Jovanovic I, Zivkovic M, Jesic S, Stankovic A. Non-coding RNA and cholesteatoma. *Laryngoscope Investigative Otolaryngology.* 2022;1-7. doi:10.1002/lio2.728