

IMPORTANCE OF COPY NUMBER ALTERATIONS OF *FGFR1* AND *c-MYC* GENES IN TRIPLE NEGATIVE BREAST CANCER

ZNAČAJ PROMENA BROJA KOPIJA *FGFR1* I *c-MYC* GENA U TROSTRUKO NEGATIVNIM KARCINOMIMA DOJKE

Milica Nedeljković¹, Nikola Tanić², Tatjana Dramićanin³, Zorka Milovanović¹, Snežana Šušnjar¹, Vedrana Milinković⁴, Ivana Vujović³, Nasta Tanić³

¹Institute of Oncology and Radiology of Serbia, Belgrade, Serbia

²Institute for Biological Research »Siniša Stanković«, University of Belgrade, Belgrade, Serbia

³Institute of Nuclear Sciences »Vinča«, University of Belgrade, Belgrade, Serbia

⁴Polyclinic Beo-lab plus, Belgrade, Serbia

Summary

Background: Triple negative breast cancer (TNBC) is characterized by aggressive clinical course and is unresponsive to anti-HER2 and endocrine therapy. TNBC is difficult to treat and is often lethal. Given the need to find new targets for therapy we explored clinicopathological significance of copy number gain of *FGFR1* and *c-MYC*. Our aim was to determine the impact of *FGFR1* and *c-MYC* copy number gain on clinical course and outcome of TNBC.

Methods: *FGFR1* and *c-MYC* gene copy number alterations were evaluated in 78 archive TNBC samples using TaqMan based quantitative real time PCR assays.

Results: 50% of samples had increased *c-MYC* copy number. *c-MYC* copy number gain was associated with TNBC in contrast to ER positive cancers. Our results showed significant correlation between *c-MYC* copy number gain and high grade of TNBCs. This suggests that *c-MYC* copy number could be an useful prognostic marker for TNBC patients. *c-MYC* copy number gain was associated with high pTNM stage as well as lobular and medullary tumor subtypes. 43% of samples had increased *FGFR1* copy number. No correlations between *FGFR1* copy number gain and clinicopathological variables were observed.

Conclusions: We identified *c-MYC* copy number gain as a prognostic marker for TNBC. Our results indicate that *c-*

Kratak sadržaj

Uvod: Trostruko negativne karcinome dojke karakteriše agresivan klinički tok i neosetljivost na endokrinu i anti-HER2 terapiju. Ovi tumori se teško leče i često su letalni. Zbog potrebe za novim tipovima terapije, ispitali smo kliničko-patološki značaj povećanja broja kopija *FGFR1* i *c-MYC* onkogeni. Cilj rada je bio da se utvrdi uticaj povećanja broja kopija *FGFR1* i *c-MYC* na klinički tok i ishod trostruko negativnog karcinome dojke.

Metode: Promene u broju kopija *FGFR1* i *c-MYC* gena određene su kvantitativnim PCR-om u realnom vremenu kod 78 arhivskih uzoraka trostruko negativnog karcinome dojke.

Rezultati: 50% ispitanih uzoraka je imalo povećan broj kopija *c-MYC*. Povećanje broja kopija *c-MYC* gena je asociirano sa trostruko negativnim karcinomima dojke u poređenju sa ER pozitivnim karcinomima. Amplifikacija *c-MYC* je asociirana sa visokim gradusom trostruko negativnih karcinoma. Iz ovog rezultata proizilazi da bi se broj kopija *c-MYC* mogao smatrati korisnim prognostičkim markerom za TNBC pacijente. Povećanje broja kopija *c-MYC* gena je asociirano i sa visokim stadijumom tumora kao i sa lobularnim i medularnim podtipom. 43% ispitanih uzoraka je imalo povećan broj kopija *FGFR1*. Nisu utvrđene nikakve korelacije između povećanja broja kopija *FGFR1* i kliničkih i histopatoloških parametara tumora.

Address for correspondence:

Milica Nedeljković
Institute of Oncology and Radiology of Serbia,
Pasterova 14, Belgrade, Serbia
e-mail address: mnedel30@yahoo.co.uk
Fax: +381 11 2685 300
Phone: +381 11 2067 210

List of abbreviations: CISH, chromogenic in situ hybridization; *c-MYC*; CNA, copy number alterations; DFI, disease free interval; ER, estrogen receptor; FFPE, formalin-fixed, paraffin-embedded; *FGFR1*, fibroblast growth factor receptor 1; IHC, immunohistochemistry; HER2, human epidermal growth factor receptor 2; OS, overall survival; PR, progesterone receptor; qPCR, quantitative real time PCR; TNBC, triple negative breast cancer

MYC may contribute to TNBC progression. We observed no significant association between *c-MYC* and/or *FGFR1* copy number status and patient survival.

Keywords: *c-MYC*; copy number gain; *FGFR1*; triple negative breast cancer

Introduction

Triple negative breast cancer (TNBC) is defined by the lack of expression of estrogen (ER), progesterone (PR) and human epidermal growth factor receptor 2 (HER2). It accounts for 10 to 20% of all invasive breast cancer cases (1). TNBC is a highly heterogeneous disease and is usually an invasive ductal carcinoma of no special type with a high histological grade and mitotic index (1). It is characterized by poor prognosis and aggressive clinical course (2–4). Currently the only systemic therapy available for TNBC is conventional cytotoxic chemotherapy but its effects seem to be insufficient (5, 6). Identifying molecular targets and devising new therapeutics for these targets is an ongoing effort and imperative for the development of a successful therapy for TNBC.

Fibroblast growth factor receptor 1 (*FGFR1*) belongs to the fibroblast growth factor receptor family of transmembrane receptor tyrosine kinases. It has an important role in many cellular processes such as cell differentiation, proliferation, migration and apoptosis (7). Aberrant *FGFR* signaling has been associated with cancerogenesis in several human cancers thus making it a potential therapeutic target (7). Gene amplification is one of the most commonly identified *FGFR1* aberrations in breast cancer. *FGFR1* amplification was shown to be associated with invasive breast cancer suggesting it could influence breast cancer progression by contributing to the invasive transition processes (8). While *FGFR1* amplification was associated with poor prognosis in ER positive breast cancer (9), its role in TNBC is far less clear. To date there have been only a few reports of *FGFR1* alternations and their impact on TNBC progression and prognosis.

c-MYC protein is a transcription factor that serves as a key regulator of most aspects of cellular function including metabolism, replication, growth, differentiation and cell death (10). *c-MYC* overexpression and gene amplification have been detected in a majority of human cancers including breast cancer. *c-MYC* expression and signaling were found to be elevated in TNBCs compared to hormone receptor positive cancers and linked to poor prognosis (11). *c-MYC* amplification was shown to be preferentially associated with invasive zones of breast cancer (12) and several studies have indicated that *c-MYC*

Zaključak: Utvrdili smo da je povećanje broja kopija *c-MYC* prognostički marker za trostruko negativne karcinome dojke. Naši rezultati pokazuju da *c-MYC* može da doprinese progresiji trostruko negativnih tumora. Nije opažena statistički značajna asocijacija između broja kopija *c-MYC* i/ili *FGFR1* onkogeni i preživljavanja pacijenata.

Ključne reči: *c-MYC*; *FGFR1*; povećanje broja kopija gena; trostruko negativan karcinom dojke

may play an important role in aggressive breast cancers with poor prognosis (13, 14).

The main goals of this study were to evaluate the rates and prognostic significance of *FGFR1* and *c-MYC* copy number alterations (CNA) in TNBC.

Materials and Methods

Samples

Seventy eight breast carcinoma samples obtained from the Institute for Oncology and Radiology of Serbia that were confirmed negative for ER, PR and HER-2 were included in this study. ER, PR and HER-2 expression were evaluated using commercial semi quantitative immunohistochemistry (IHC) assays, Dako, according to the manufacturer recommended procedure. The scoring system included the percentage of stained cells on a score from 0 to 5, and the intensity of their staining on a score from 0 to 3. Cases with the overall IHC score <4 were considered negative for ER and PR expression (15). An overall score of 0 or 1+ for IHC staining of HER2 expression was regarded as negative. Score of 2+ was considered equivocal and for these cases HER2 negative status was confirmed by chromogenic in situ hybridization (CISH) (16).

All tumor samples and their corresponding normal tissue were formalin-fixed, paraffin-embedded (FFPE). All relevant histopathologic and clinical parameters (age, tumor type, pN stage, pT stage, pTNM stage, Nottingham combined histologic grade, disease free survival, overall survival) were retrieved from patient's medical records.

This study was approved by the Institute for Oncology and Radiology of Serbia ethics committee number 4321-01 and carried out in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki, the International Ethical Guidelines for Biomedical Research Involving Human Subjects (CIOMS), Geneva 1993, and the Guidelines for Good Clinical Practice CPMP/ICH/135/95), September 1997.

DNA extraction

Genomic DNA was extracted from all samples using Kapa Biosystems Express Extract Kit (KK7151, Kapa Biosystems, Wilmington, MA, USA) according to the manufacturers recommended protocol. The quality of the extracted DNA was verified by agarose gel electrophoresis. Concentrations and purity were assessed spectrophotometrically using A260/A280 absorbance ratios (NanoDrop Technologies, Wilmington, DE, USA). Extracted and purified DNA was stored at +4 °C until further analyses.

Copy number analysis by quantitative real time PCR

Copy number analyses of *FGFR1* and *c-MYC* genes were done by quantitative real time PCR (qPCR) using TaqMan based assays. *c-MYC* assay included highly specific forward and reverse primers as well as a 6-Fam-TAMRA labelled probe: F5'GGAC-GACGAGACCTTCATCAA-3', R5'-CCAGCTTCTCT-GAGACGAGCTT-3', TaqMan Probe 6-FAM-5'-AGAAGCCGCTCCACATACAGTCCTGG-3'-TAMRA. *FGFR1* gene copy number was evaluated using Hs00237051_cn TaqMan assay (Applied Biosystems, Foster City, CA, USA). RNase-P was used as a reference gene (4403326, Applied Biosystems). Calibration was done using DNA isolated from normal breast tissue.

Each sample was prepared in duplicate while normal DNA controls were prepared in triplicate. Total reaction volume was 15 µL. For *c-MYC*, reaction contained primers/probe ratio of 3:1 (0.1 mmol/L probe: 0.3 mmol/L primers), 1x TaqMan Master Mix and 40 ng of DNA. For *FGFR1* or RNase-P reaction contained 1x TaqMan Master Mix, 1x TaqMan Copy Number Assay for *FGFR1* or RNase-P gene and 40 ng of DNA. Each reaction contained two normal DNA controls that were used as calibrators. PCR reactions were carried out in the ABI Prism 7500 Sequence Detection System at 95 °C for 10 minutes, followed by 40 cycles at 95 °C for 15 seconds, and 60 °C for 1 minute. A mean Ct value of each duplicate was used for relative quantitation of gene copy number according to the Livak ($2^{-\Delta\Delta CT}$) method. Each run included a no-template control. The obtained results were analysed by RQ Study Add ON software for 7500 v 1.3 SDS instrument with a confidence level of 95% ($p < 0.05$).

FGFR1 and *c-MYC* copy number were classified as gain if average copy number ratio estimate given by qPCR was ≥ 2.0 .

Statistical analysis

Data analysis was performed with GraphPad Prism 5 software (GraphPad Software, Inc. CA). The

correlations between clinicopathologic parameters and CNA of *c-MYC* and *FGFR1* genes were analysed using Fisher's exact test or the Chi-square test, depending on test conditions. Survival analyses were performed using Kaplan & Meier product-limit method and the log rank test was used to determine the significance of the difference between survival curves. Overall survival (OS) was calculated from the day after surgery to the last follow-up examination or death of the patient and disease free survival (DFS) was defined as time from the day after surgery to first locoregional recurrence, distant recurrence or contralateral disease. Statistical differences were considered significant when p value was < 0.05 .

Results

Copy number alternations of *c-MYC* and *FGFR1* oncogenes

We analyzed 78 TNBC samples out of which 34 had increased *FGFR1* copy number (43%) and 39 (50%) had increased *c-MYC* copy number. Comparing this findings with our previous results on receptor positive breast cancer samples (17) it is evident that the triple negative cohort has a significantly higher incidence of copy number gain for *c-MYC* ($p < 0.005$).

We then correlated copy number gain of examined genes with the clinicopathologic parameters of TNBC (Table I). Copy number gain of *c-MYC* oncogene was significantly associated with high histological grade ($p = 0.008$) and high pTNM stage (Table I). Patients with lobular breast cancer were represented more in the group with increased *c-MYC* copy number ($p = 0.014$) and *c-MYC* copy number gain group tended to have a higher representation of medullary breast cancers ($p = 0.06$). Surprisingly, the group with no *FGFR1* copy number gain had a higher percentage of pTNM stage 4 tumors (Table I). *FGFR1* copy number gain was not associated with any other clinicopathologic variable. Neither of the analyzed genes had any significant influence on patient OS and DFS (Figure 1).

Next, we analysed whether there were co-alterations between *FGFR1* and *c-MYC* oncogenes. 24 (31%) of samples had increased both *FGFR1* and *c-MYC* copy number, 25 (32%) had copy number gain of one gene and 29 (37%) had no copy number gain for either gene. Simultaneous copy number gain of both genes was significantly associated with high histological grade ($p = 0.039$) and pTNM stage III (Table I). Patients with no copy number gain for either gene had a higher proportion of pTNM stage I tumors compared to other groups while patients with copy number gain of one gene had higher proportion of pTNM stage IV tumors (Table I). No correlation with patient survival was obtained (data not shown).

Table I Clinicopathological correlation according to *c-MYC* and/or *FGFR1* copy number alterations in TNBC.

Parameters	Total (np=78)	c-MYC np (%)		FGFR1 np (%)		c-MYC and FGFR1 np (%)		
		Gain (np=39)	p value	Gain (np=34)	p value	Both gain (np=24)	One gain (np=25)	p value
Tumor type								
Ductal	46 (59)	18 (46)	p^A 0.014	18 (53)	p ^A 1.000	11 (46)	14 (56)	p ^A 0.220
Lobular	14 (18)	11 (28)	p ^B 0.064	6 (18)	p ^B 0.129	6 (25)	5 (20)	p ^B 0.071
Medullary	14 (18)	10 (26)	p ^C 0.283	9 (26)	p ^C 1.000	7 (29)	5 (20)	p ^C 0.438
Other*	4 (5)	0 (0)	p ^D 1.000	1 (3)	p ^D 0.449	0 (0)	1 (4)	p ^D 0.871
			p^E 0.011		p ^E 1.000			p ^E 0.105
			p^F 0.023		p ^F 0.274			p^F 0.044
Stage								
I	10 (13)	1 (2)	p¹ 0.031	3 (9)	p ¹ 0.292	1 (4)	2 (8)	p ¹ 0.106
II	38 (49)	19 (49)	p² 0.002	20 (59)	p ² 0.198	14 (59)	11 (44)	p² 0.021
III	11 (14)	9 (23)	p³ 0.044	7 (20)	p ³ 0.665	7 (29)	2 (8)	p ³ 0.204
IV	19 (24)	10 (26)	p ⁴ 0.087	4 (12)	p ⁴ 0.732	2 (8)	10 (40)	p ⁴ 0.284
			p ⁵ 1.000		p⁵ 0.027			p ⁵ 0.079
			p ⁶ 0.139		p⁶ 0.046			p⁶ 0.009
pT stage								
T1 and T2	68 (87)	34 (87)	1.000	28 (82)	0.317	21 (87)	20 (80)	0.356
T3 and T4	10 (13)	5 (13)		6 (18)		3 (13)	5 (20)	
Nodal status								
Negative	32 (41)	20 (51)	1.000	21 (64)	0.116	14 (59)	12 (48)	0.708
Positive	46 (59)	19 (49)		13 (36)		10 (41)	13 (52)	
Histologic grade								
I and II	52 (67)	20 (51)	0.008	20 (59)	0.237	12 (50)	16 (64)	0.039
III	26 (33)	19 (49)		14 (41)		12 (50)	9 (36)	

Abbreviations: np, number of patients per group; * – tubular, mucinous and other rare carcinoma types

p^A – statistical significance between ductal and lobular tumors; p^B – statistical significance between ductal and medullary tumors; p^C – statistical significance between ductal and other tumors; p^D – statistical significance between lobular and medullary tumors; p^E – statistical significance between lobular and other tumors; p^F – statistical significance between medullary and other tumors

p¹ – statistical significance between stages I and II; p² – statistical significance between stages I and III; p³ – statistical significance between stages I and IV; p⁴ – statistical significance between stages II and III; p⁵ – statistical significance between stages II and IV; p⁶ – statistical significance between stages III and IV

Bold indicates statistically significant values, p < 0.05.

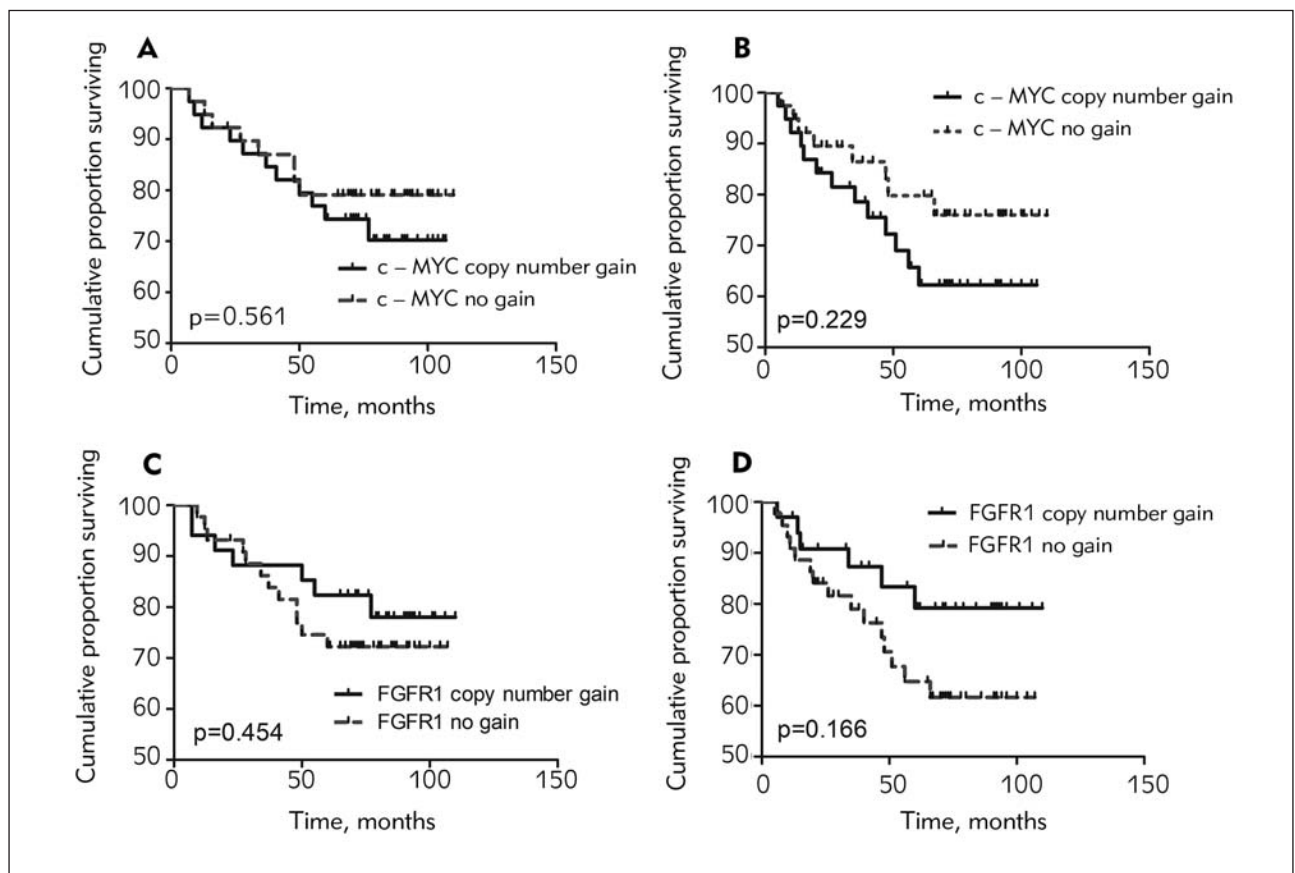


Figure 1 Kaplan–Meier survival curves. Impact of *c-MYC* and *FGFR1* copy number on patient’s overall survival and disease free survival. A *c-MYC* copy number gain had no influence on patient overall survival. B *c-MYC* copy number gain had no influence on patient disease free survival. C *FGFR1* copy number gain had no influence on patient overall survival. D *FGFR1* copy number gain had no influence on patient disease free survival

Discussion

Gene copy number gain is an important mechanism of oncogene activation in cancer however the prognostic significance of *FGFR1* CNA in TNBC remains unclear. Our study showed that *FGFR1* gene copy number was increased in 43% of examined samples which is a higher frequency than previously reported by Lee et al. (18). However, Lee et al. (18) examined *FGFR1* amplification in TNBC using fluorescence in situ hybridization (FISH). A recent study compared qPCR with FISH for assessing gene copy number (19). qPCR exhibited excellent correlation with FISH at detecting copy number gain at 8 or more copies. In the range from 2 to 6 copy number gain as detected by qPCR, no copy number gains were found using FISH. We would argue that qPCR is a more sensitive method for detecting lower levels of copy number gain. In our cohort the majority of samples with gene amplification had a low-grade copy number increase. Therefore it is very likely that the higher frequency of *FGFR1* copy number gain detected in our study is due to the higher sensitivity of qPCR. In the same study Park et al. (19) demonstrated that FFPE tissue showed lower levels of copy number gain compared to frozen

tissue therefore qPCR would be ideally suited for assessing gene copy number gain in FFPE tissue.

FGFR1 expression has impact on overall survival in TNBC but its role in this disease is still controversial (20). Literature data is scarce and offers conflicting results. Cheng et al. (20) found that *FGFR1* expression in TNBCs was independently predictive for OS with cases with high *FGFR1* immunostaining having the worst prognosis. Another study by Lee et al. (18) showed no correlation between *FGFR1* expression and patient survival. The two studies differed in immunostaining threshold used which highlights the difficulty of comparing data from various studies. Underlying mechanisms of *FGFR1* overexpression in TNBC are not well understood. While several studies reported high level of correlation between *FGFR1* protein overexpression and gene amplification, (21, 22), a number of studies observed low protein expression level in *FGFR1* amplified tumors (23, 24). We found no significant association between *FGFR1* copy number gain and poor prognosis in TNBC or any of the clinicopathologic parameters. Our results support the notion that *FGFR1* copy number status may not be an informative independent prognostic factor for TNBC.

c-MYC is frequently deregulated in breast cancer and is thought to contribute to breast cancer progression and poor prognosis. However, the detected frequency of *c-MYC* amplification and its prognostic significance have been inconsistent (25, 26) with the reported frequency ranging from 1 to 94% (10). We have detected *c-MYC* copy number gain in 50% of TNBC samples, a frequency significantly higher than in receptor positive breast cancers we previously examined (17). Dillon et al. (27) detected that 75% of TNBCs and 89% of basal-like tumors had *c-MYC* amplification. However, this study analyzed only 20 TNBC samples. Here we have analyzed a much larger cohort and demonstrated that *c-MYC* copy number gain is a frequent event in TNBC. High frequency of *c-MYC* overexpression and copy number gain observed in TNBC, indicates that *c-MYC* deregulation could be important for TNBC progression.

We have found that *c-MYC* copy number gain was associated with high grade TNBC. Determining whether *c-MYC* copy number gain is present may help identify patients with a greater risk of developing high grade TNBC. Therefore, *c-MYC* could be considered as a prognostic marker of tumour progression.

Copy number gain of *c-MYC* was significantly associated with stage III of TNBC. These results are in compliance with previous studies that found *c-MYC* amplification to be associated with invasive zones of breast cancer. These results further confirm that *c-MYC* copy number gain could be a prognostic marker of tumour progression in TNBC.

Our result that medullary tumors were associated with *c-MYC* copy number gain is consistent with a previous study (28). This suggests that medullary tumors have a distinct biology conducive to *c-MYC* amplification. Our finding that lobular tumors were associated with *c-MYC* copy number gain differs from a study by Green et al. (29) which found *c-MYC* expression to be more frequent in non-lobular tumors. However, in the study by Green et al. (29) only 15% of samples were TNBC. Our results support the notion that triple negative lobular carcinoma is genetically distinctive from non-triple negative lobular carcinoma. Previous studies have shown that these two types of lobular carcinoma differ in clinicopathologic and IHC characteristics (30).

It has been shown that *c-MYC* overexpression is associated with TNBC. Understanding the underlying mechanisms of *c-MYC* expression may open new approaches for therapy of TNBC. One study showed that CDK inhibition effectively induced tumor regression in TNBC tumors that exhibit elevated *c-MYC*

expression (11). *c-MYC* could represent a promising new target for TNBC therapy.

We have found a high level of concordance between *FGFR1* and *c-MYC* copy number gain as 24 (31%) of samples had increased both *FGFR1* and *c-MYC* copy number. Simultaneous copy number gain of both genes was associated with high grade and high pTNM stage. However, this might be due to the association of *c-MYC* gain with these parameters. Simultaneous absence of copy number gain for both genes was associated with pTNM stage I which further supports the notion that CNA of these genes occurs at a later phase of tumor progression. One previous study identified frequent coamplification of *c-MYC* and 8p11-p12 chromosomal region, where *FGFR1* is located, in invasive breast cancers (31). Additionally, evidence exists that links *c-MYC* expression with more favorable response to FGFR inhibiting therapy in lung cancers that exhibit *FGFR1* amplification (32). Therefore, examining *FGFR1* and *c-MYC* copy number and expression together may help determine which patients would benefit the most from FGFR inhibiting therapy.

In conclusion, copy number gain of *c-MYC* is associated with high grade and high stage TNBC as well as lobular and medullary tumor subtypes. *FGFR1* gene copy number has low prognostic implication for TNBC. High level of concordance in *c-MYC/FGFR1* copy number gain was detected in this cohort. Simultaneous copy number gain of both genes was significantly associated with high histological grade and pTNM stage of TNBC. Though we found no significant implications for patient outcome, a subset of TNBC harbor copy number gain of *c-MYC* and *FGFR1* which could be of interest for TNBC therapy. Patients with no copy number gain for either gene had a higher proportion of pTNM stage I tumors confirming the predictive importance of these genes amplification for tumor progression.

Acknowledgments.

Funding. This research was supported by the Ministry of Education, Science and Technological Development, Republic of Serbia, grant number III41031 and grant number ON173049.

Conflict of interest statement

The authors state that they have no conflicts of interest regarding the publication of this article.

References

- Foulkes WD, Smith IE, Reis-Filho JS. Triple-Negative Breast Cancer. *N Engl J Med* 2010; 1938–48.
- Collinson P. Laboratory medicine is faced with the evolution of medical practice. *J Med Biochem* 2017; 36: 211–5.
- Albergaria A, Ricardo S, Milanezi F, Carneiro V, Amendoeira I, Vieira D, et al. Nottingham Prognostic Index in triple-negative breast cancer: a reliable prognostic tool? *BMC Cancer* 2011; 11(1): 299.
- Dent R, Trudeau M, Pritchard KI, Hanna WM, Kahn HK, Sawka CA, et al. Triple-negative breast cancer: Clinical features and patterns of recurrence. *Clin Cancer Res* 2007; 13(15): 4429–34.
- Liedtke C, Mazouni C, Hess KR, André F, Tordai A, Mejia JA, et al. Response to neoadjuvant therapy and long-term survival in patients with triple-negative breast cancer. *J Clin Oncol* 2008; 26(8): 1275–81.
- Peddi PF, Ellis MJ, Ma C. Molecular basis of triple negative breast cancer and implications for therapy. *Int J Breast Cancer* 2012; 217185.
- Haugsten EM, Wiedlocha A, Olsnes S, Wesche J. Roles of fibroblast growth factor receptors in carcinogenesis. *Mol Cancer Res* 2010; 8(11): 1439–52.
- Jang M, Kim E, Choi Y, Lee H, Kim Y, Kim J, et al. FGFR1 is amplified during the progression of in situ to invasive breast carcinoma. *Breast Cancer Res* 2012; 14(4): R115.
- Elbauomy Elsheikh S, Green AR, Lambros MBK, Turner NC, Grainge MJ, Powe D, et al. FGFR1 amplification in breast carcinomas: a chromogenic in situ hybridisation analysis. *Breast Cancer Res* 2007; 9(2): R23.
- Liao DJ, Dickson RB. c-Myc in breast cancer. *Endocr Relat Cancer* 2000; 7(3): 143–64.
- Horiuchi D, Kusdra L, Huskey NE, Chandriani S, Lenburg ME, Gonzalez-Angulo AM, et al. MYC pathway activation in triple-negative breast cancer is synthetic lethal with CDK inhibition. *J Exp Med* 2012; 209(4): 679–96.
- Corzo C, Corominas JM, Tusquets I, Salido M, Bellet M, Fabregat X, et al. The MYC oncogene in breast cancer progression: from benign epithelium to invasive carcinoma. *Cancer Genet Cytogenet* 2006; 165(2): 151–6.
- Aulmann S. c-myc Amplifications in primary breast carcinomas and their local recurrences. *J Clin Pathol* 2006; 59(4): 424–8.
- Blancato J, Singh B, Liu A, Liao DJ, Dickson RB. Correlation of amplification and overexpression of the c-myc oncogene in high-grade breast cancer: FISH, in situ hybridisation and immunohistochemical analyses. *Br J Cancer* 2004; 90(8): 1612–9.
- Leake R. Immunohistochemical detection of steroid receptors in breast cancer: a working protocol. *J Clin Pathol* 2000; 53(8): 634–5.
- Wolff AC, Hammond MEH, Schwartz JN, Hagerty KL, Allred DC, Cote RJ, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for human epidermal growth factor receptor 2 testing in breast cancer. *J Clin Oncol* 2007; 25(1): 118–45.
- Tanić N, Milinković V, Dramićanin T, Nedeljković M, Stanković T, Milovanović Z, et al. Amplification of Cyclin D1, C-MYC And EGFR Oncogenes in Tumour Samples of Breast Cancer Patients. *J Med Biochem* 2013; 32(4): 1–8.
- Lee HJ, Seo AN, Park SY, Kim JY, Park JY, Yu JH, et al. Low Prognostic Implication of Fibroblast Growth Factor Family Activation in Triple-negative Breast Cancer Subsets. *Ann Surg Oncol* 2014; 1561–8.
- Park YS, Na Y-S, Ryu M-H, Lee C-W, Park HJ, Lee J-K, et al. FGFR2 Assessment in Gastric Cancer Using Quantitative Real-Time Polymerase Chain Reaction, Fluorescent In Situ Hybridization, and Immunohistochemistry. *Am J Clin Pathol* 2015; 143(6): 865–72.
- Cheng CL, Thike AA, Tan SYJ, Chua PJ, Bay BH, Tan PH. Expression of FGFR1 is an independent prognostic factor in triple-negative breast cancer. *Breast Cancer Res Treat* 2015; 151(1): 99–111.
- Turner N, Pearson A, Sharpe R, Lambros M, Geyer F, Lopez-Garcia MA, et al. FGFR1 amplification drives endocrine therapy resistance and is a therapeutic target in breast cancer. *Cancer Res* 2010; 70(5): 2085–94.
- Tomiguchi M, Yamamoto Y, Yamamoto-ibusuki M, Gotoyamaguchi L, Fujiki Y, Fujiwara S, et al. Fibroblast growth factor receptor-1 protein expression is associated with prognosis in estrogen receptor-positive / human epidermal growth factor receptor-2-negative primary breast cancer. *Cancer Sci* 2016; 107(4): 491–8.
- Sousa V, Reis D, Silva M, Alarcão AM, Ladeirinha AF, d'Aguiar MJ, et al. Amplification of FGFR1 gene and expression of FGFR1 protein is found in different histological types of lung carcinoma. *Virchows Arch* 2016; 469(2): 173–82.
- Seo AN, Jin Y, Lee HJ, Sun PL, Kim H, Jheon S, et al. FGFR1 amplification is associated with poor prognosis and smoking in non-small-cell lung cancer. *Virchows Arch* 2014; 465(5): 547–58.
- Xu J, Chen Y, Olopade OI. MYC and Breast Cancer. *Genes Cancer* 2010; 1(6): 629–40.
- Deming SL, Nass SJ, Dickson RB, Trock BJ. C-myc amplification in breast cancer: a meta-analysis of its occurrence and prognostic relevance. *Br J Cancer* 2000; 83(12): 1688–95.
- Dillon JL, Mockus SM, Ananda G, Spotlow V, Wells WA, Tsongalis GJ, et al. Somatic gene mutation analysis of triple negative breast cancers. *Breast* 2016; 29: 202–7.
- Al-kuraya K, Schraml P, Torhorst J, Tapia C, Zaharieva B, Novotny H, et al. Prognostic Relevance of Gene Amplifications and Coamplifications in Breast Cancer. *Cancer Res* 2004; (33): 8534–40.
- Green AR, Aleskandarany MA, Agarwal D, Elsheikh S, Nolan CC, Diez-Rodriguez M, et al. MYC functions are specific in biological subtypes of breast cancer and confers resistance to endocrine therapy in luminal tumours. *Br J Cancer* 2016; 114(8): 917–28.

30. Koo JS, Jung W. Clinicopathologic and immunohistochemical characteristics of triple negative invasive lobular carcinoma. *Yonsei Med J* 2011; 52(1): 89–97.
31. Parris TZ, Kovács A, Hajizadeh S, Nemes S, Semaan M, Levin M, et al. Frequent MYC coamplification and DNA hypomethylation of multiple genes on 8q in 8p11-p12-amplified breast carcinomas. *Oncogenesis* 2014; 3(3): e95.
32. Malchers F, Dietlein F, Schöttle J, Lu X, Nogova L, Albus K, et al. Cell-autonomous and non-cell-autonomous mechanisms of transformation by amplified *FGFR1* in lung cancer. *Cancer Discov* 2014; 4(2): 246–57.

Received: February 13, 2018

Accepted: March 18, 2018