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BINDING OF 17-SUBSTITUTED 16-NITRILE 16,17-SECOESTRANE COMPOUNDS TO ESTROGEN RECEPTORS - *IN VITRO* AND *IN SILICO* STUDY

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Abstract

About 75% of breast cancers express estrogen receptors (ERs), which is a good base for an efficient endocrine therapy. This gives the opportunity for the treatment of patients with antiestrogens, compounds that bind to the ERs and thus compete to estradiol (E2), preventing its action in progression of estrogen-depending cancers.

Here we present results of testing the effect of the modified steroids, namely 17-substituted 16-nitrile 16,17-secoestrane compounds on the E2-ER complex forming, its stability, nuclear translocation and binding to DNA. Almost all compounds in moderate to high rate induced lower forming of this complex, destabilizing it – they increased K_d of this complex and decreased number of binding sites. Complex formed in the presence of some test secosteroids could pass to the nucleus, while other compounds inhibited translocation. In the presence of some compounds binding of the formed complex E2-ER to DNA was noticed.

Docking followed molecular dynamics simulation was performed to reveal binding mode of E2 to ER in the presence of test secosteroids. Amino acids important for binding process and complex stabilization were detected. Analysis of the simulation data allowed identifying key amino acids and type of binding of the secoestrane compounds, important for high affinity binding of the steroidal compounds.

Keywords: ER, secoestrane compounds, docking, molecular dynamics

1. Introduction

17β-Estradiol (E2), the main circulating estrogen hormone, regulates many physiological functions, where many of them are specific for female reproductive tissues. The effects of E2 on human cells are mediated by the estrogen receptors ER α and ER β and complex transcription machinery. The ERs express similar, still different, regulatory potentials of cellular growth, differentiation and death. According to that, changes in E2 signaling *via* E2-ER could be important factors in the initiation and progression of malignancies, including cancers of reproductive tissues [1].

Many research studies are directed towards better understanding of the mechanisms which encompasses ERs, in order to get information necessary for discovery of novel and better drugs

for treatment of estrogen hormone dependent diseases, with substantial impacts on the systemic management of target tissue [2, 3].

There are two main strategies in the biology-driven medicinal chemistry of the drug development for the treatment of hormone-dependent diseases: A. development of compounds that inhibit enzymes that are essential for biosynthesis of active steroid hormones (e.g., aromatase for the treatment of estrogen-dependent breast cancer) and B. development of compounds that compete with the appropriate receptor proteins (e.g. estrogen receptor antagonists, such as 4-hydroxytamoxifen), as presented at Figure 1.



Fig. 1. Different strategies in the treatment of estrogen-dependent diseases: A. Lowering of biosynthesis of active estrogen hormones by inhibition of aromatase (CYP19) activity, and B. Lowering of the estradiol effect by blocking of E2-ER complex forming.

Binding of natural ligand or antagonist (steroidal or non-steroidal) causes different changes in the tertiary structure of the receptor, which allows ER signaling pathway continuation or suspension, as discovered in the studies of the complexes of ER with E2 and ER antagonist 4hydroxytamoxifen [4, 5].

Modified estradiol-based steroids, including substituted, homo-, epi- and secoestranes, proved to be good partial or full antiestrogens (selective estrogen receptor modulators, SERMs, or down regulators, SERDs). Among them, D ring-secosteroids were of particular interest, since the modification of the rigid tetracyclic steroidal carbon skeleton by cleavage of the internal C-C bond provides more flexible compounds with new biological properties.

Based on known pharmacological effects of some secoestrane compounds [6, 7], binding of 16,17-secoestrane compounds was tested independently in *in vitro* and *in silico* experiments and results are compoared.

2. Results and discussion

The effect of secosteroids onto the estradiol binding to the estrogen receptors is measured *in vitro* by known procedure [7], by measuring binding parameters, namely dissociation constant of E2-ER complex and receptor number, as well as the forming, stability, nuclear translocation and binding to DNA of the complex formed. Almost all 17-substituted 16,17-secoestrane compounds inhibited E2-ER complex forming, which was obvious from increasing of K_d of this complex and decreasing of binding sites number in the presence of secosteroids. Complex formed in the presence of some test secosteroids in some extent could pass to the nucleus, while other compounds inhibited translocation. In the presence of some compounds binding of the formed complex E2-ER to DNA was noticed.

Docking of ER α ligands was performed using of AutoDock Vina software (v.1.1.2) with next parameters: exhaustiveness: 256; energy range: 4 kcal/mol; number of binding modes: 20. Box dimensions and coordinates of the "docking box" for the tested compounds were picked according to their size. After docking all hits were ranked according to their RMSD towards similar known ligands, presented in corresponding structures from Protein Data Bank and according to values of AutoDock Vina scoring function. Spatial structures of steroidal derivatives were built from their 2D structures using MolView service. Structures were minimized in UCSF Chimera software before docking procedure. Molecular dynamics (MD) simulations were performed in explicit solvent (water, octahedron cell, TIP3P model). Experiment included next steps: minimization of the solvent (2000 steps), minimization of the whole system (2000 steps), heating of the system (NVT ensemble, 1 ns, temperature range: 0-298.15 K), equilibration (NPT ensemble, 1 ns), production (NPT ensemble, 50 ns).

In silico screening of binding affinity of the modified steroids toward ER α showed that interaction energy for the tested ligands is comparable with consequent value, calculated for the estradiol. Analysis of MD trajectories showed that tested compounds do not act dramatically on the receptor structure. It was found that radius of gyration (Rg) for all complexes quite similar and does not exceed 19,5 Å. Analysis of RMSF curves states that main differences between ligand-receptor complexes are located in coils, while amino acids forming binding pocket (residues 344-351, 386-394, 422-430, 518-525) are relatively stable during 50 ns MD simulation. RMSD for C_a atoms of the receptor also does not exceed 4.5 Å during simulation.

Modified steroids interact with key points of the flexible hydrophobic ligand binding pocket (LBP) of estrogen receptor α (ER α), occupying the remaining large volume of the LBP equivalent to rings of the steroids

3. Conclusions

In silico studies (docking and molecular dynamics simulation) were performed to reveal binding mode of E2 to ER in the presence of test secosteroids and amino acids important for binding to ER, as well as E2-ER complex stabilization. Simulation data directed identification of the key amino acids and type of binding of the secoestrane compounds, which explain high affinity binding of steroidal compounds.

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